



Synthesis and biological activity of N-substituted aminocarbonyl-1,3-dioxolanes as VLA-4 antagonists

Abdul Rehman^{a,*}, Ajay Soni^a, Keshav Naik^a, Sreeji Nair^a, Venkata P. Palle^a, Sunanda Dastidar^b, Abhijit Ray^b, M. S. Alam^c, Mohammad Salman^a, Ian A. Cliffe^a, Viswajanani Sattigeri^a

^a Department of Medicinal Chemistry, New Drug Discovery Research, Ranbaxy Research Laboratories, Plot 20, Sector 18, Gurgaon 122015, Haryana, India

^b Department of Pharmacology, New Drug Discovery Research, Ranbaxy Research Laboratories, Plot 20, Sector 18, Gurgaon 122015, Haryana, India

^c Department of Chemistry, Jamia Hamdard, New Delhi 110062, India

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ABSTRACT

A novel set of compounds with a 1,3-dioxolane ring which acts as a proline bioisostere have been successfully designed as VLA-4 receptor antagonists. Compounds (**18e**), (**28j**), and (**35g**) were shown to have high receptor affinities.

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Very late antigen-4 (VLA-4, $\alpha 4\beta 1$) is a member of the $\beta 1$ sub-family of integrins and is expressed on all circulating leukocytes except platelets.^{1,2} VLA-4 mediates both cell–cell and cell–matrix interactions by binding to cell surface ligands such as vascular cell adhesion molecule-1 (VCAM-1) and cell–matrix proteins such as fibronectin. These interactions are important for the activation, migration, and proliferation of leukocytes during normal and path-

ophysiological processes, and so the inhibition of VLA-4 is expected to be of therapeutic benefit in the treatment of a variety of inflammatory diseases.^{3,4}

VLA-4 binds to a motif containing the sequence EILDVPST within VCAM-1, and peptidomimetic inhibitors of the critical leucine–aspartic acid–valine (LDV) sequence have been reported.⁵ Non-peptidomimetic inhibitors have also been developed (Fig. 1) in

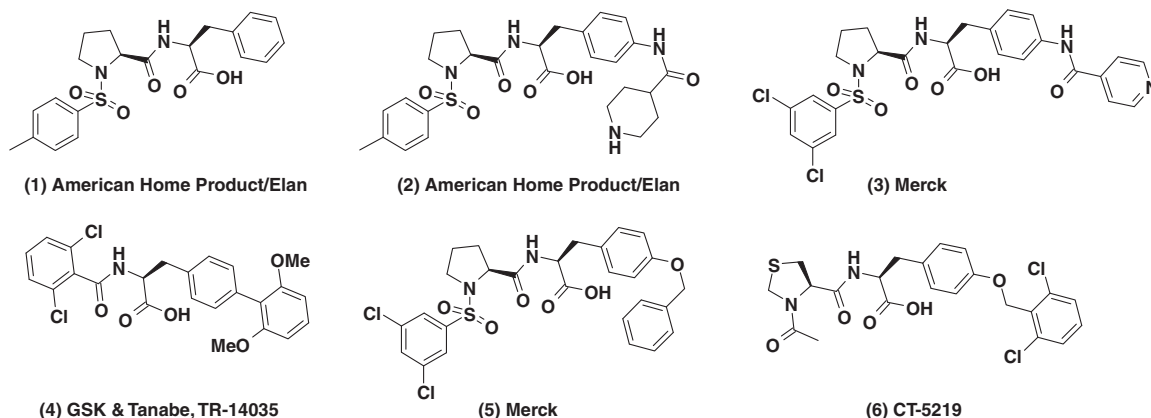
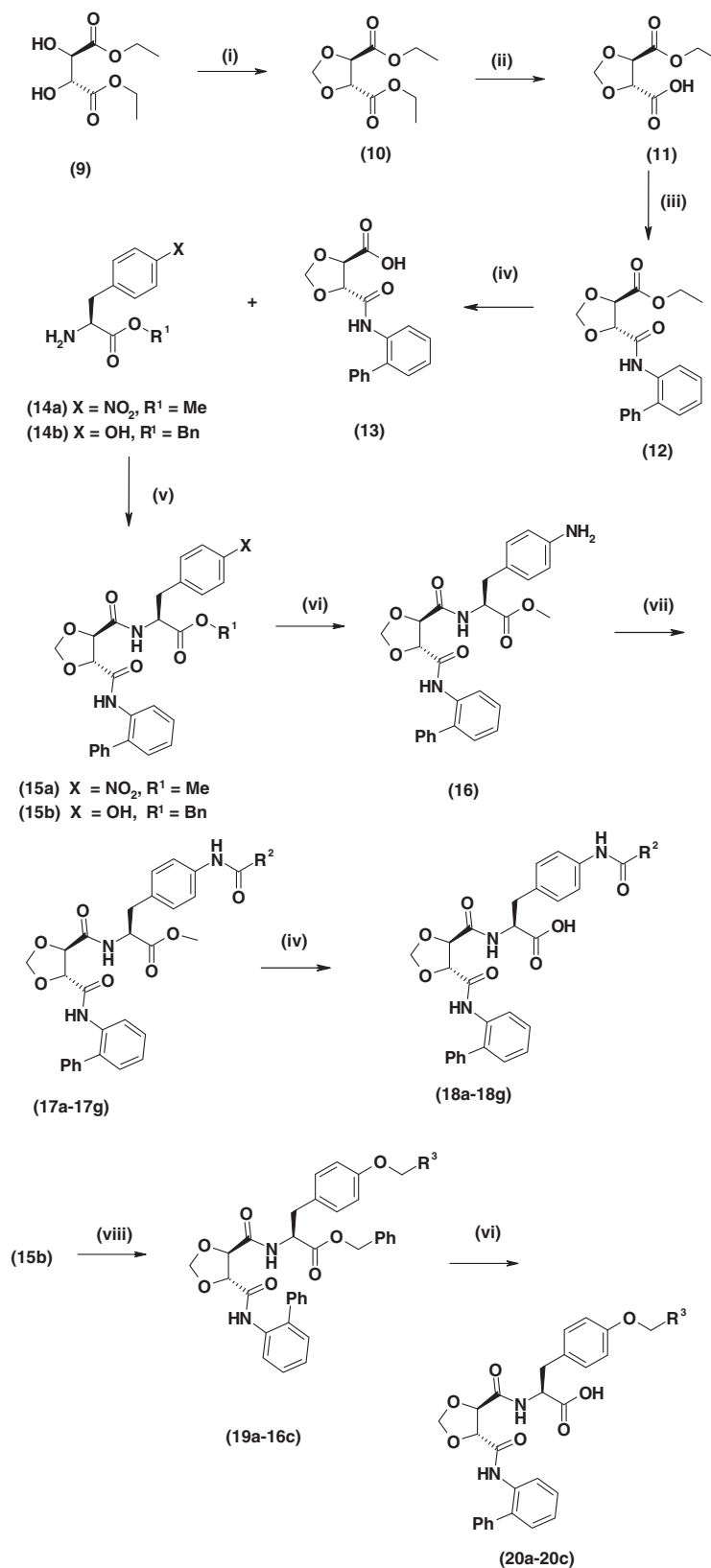


Figure 1. Potent VLA-4 inhibitors.

* Corresponding author. Tel.: +91 09999100317; fax: +91 124 2343545.

E-mail address: ab_rehman@rediffmail.com (A. Rehman).



Scheme 1. Reagents: (i) dimethoxymethane, BF₃·EtO, isopropyl acetate, 100%; (ii) KOH, EtOH, 56%; (iii) 2-phenylaniline, EDC, NMM, HOBt, DMF, 67%; (iv) LiOH·H₂O, THF:MeOH:H₂O, 50–95%; (v) EDC, NMM, HOBt, DMF, 73%; (vi) 10% Pd/C, MeOH, 87%; (vii) R²-COOH, EDC, NMM, HOBt, DMF, 45–75%; (viii) R³-CH₂Br, K₂CO₃, acetone, 51–80%.

which an *N*-acylphenylalanine core appears to be critical for activity. Unlike the LDV-based mimics which in general are very selective for VLA-4, *N*-acylphenylalanine derivatives are dual inhibitors

of VLA-4/VCAM and $\alpha 4\beta 7$ /MAdCAM (mucosal adhesion cell adhesion molecule) interactions. Elan has reported the SAR of compound (1) (CT-757) and shown that its oral bioavailability in rats

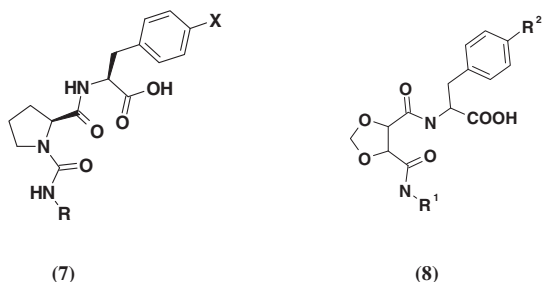


Figure 2. Proline-based and 1,3-dioxolane-based VLA-4 inhibitors.

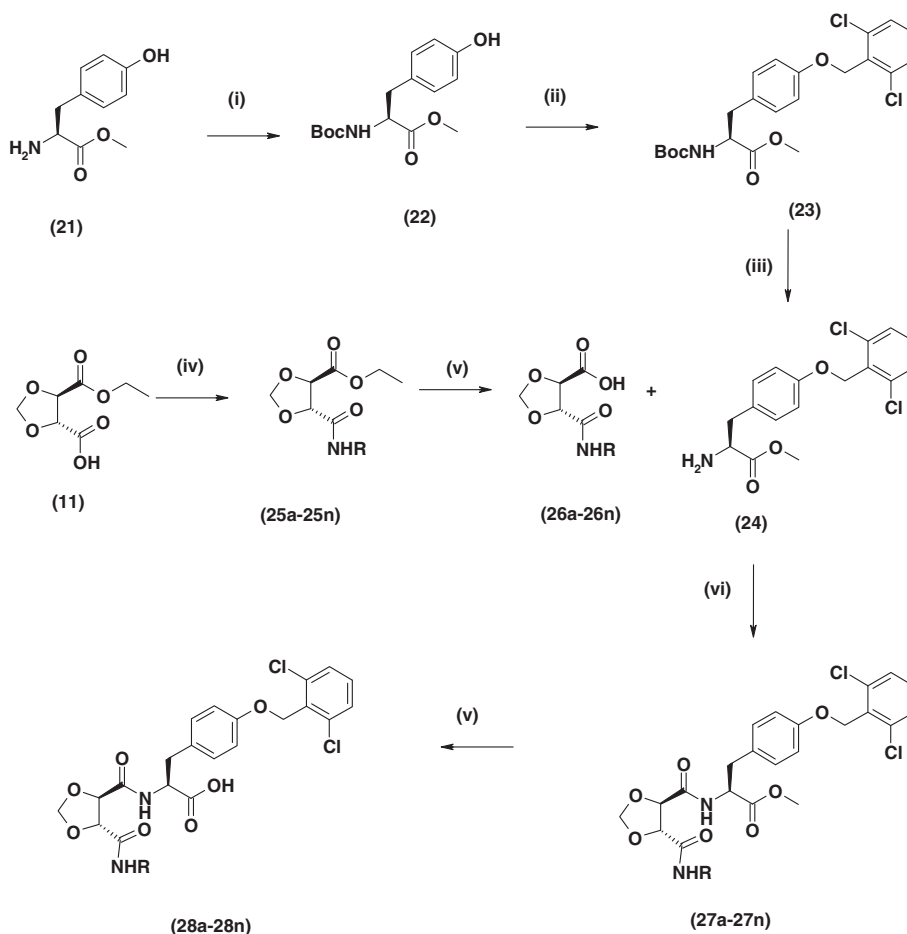
was poor (10–15%) with most of the drug being eliminated rapidly in the bile.⁶ Two potent analogs of compound (1) are the piperidine (2, IC_{50} = 40 nM) and pyridine (3, IC_{50} = 0.08 nM) which had less protein binding and lower bioavailability.^{7,8} The biphenylalanine (4) (TR-14035) and the tyrosine analog (5) have also shown potent VLA-4 antagonistic activity⁹ and Celltech has reported that the thia-proline (6, CT-5219) is a very potent VLA-4 antagonist (IC_{50} = 35 nM). This latter compound exhibited poor bioavailability in rats and sheep and showed rapid elimination, particularly in rats.^{10,11}

Literature suggests that derivatization of the phenylalanine portion of *N*-acylphenylalanine inhibitors has been a major focus of attempts to obtain optimal potency and selectivity for $\alpha_4\beta_1$. Earlier, we reported substituted proline ureas of general formula (7) as VLA-4 inhibitors (Fig. 2) in which the ureido group acted as a bioisostere of the sulfonamide group in compounds (1–3).¹² It was found that the compounds with highest binding affinities con-

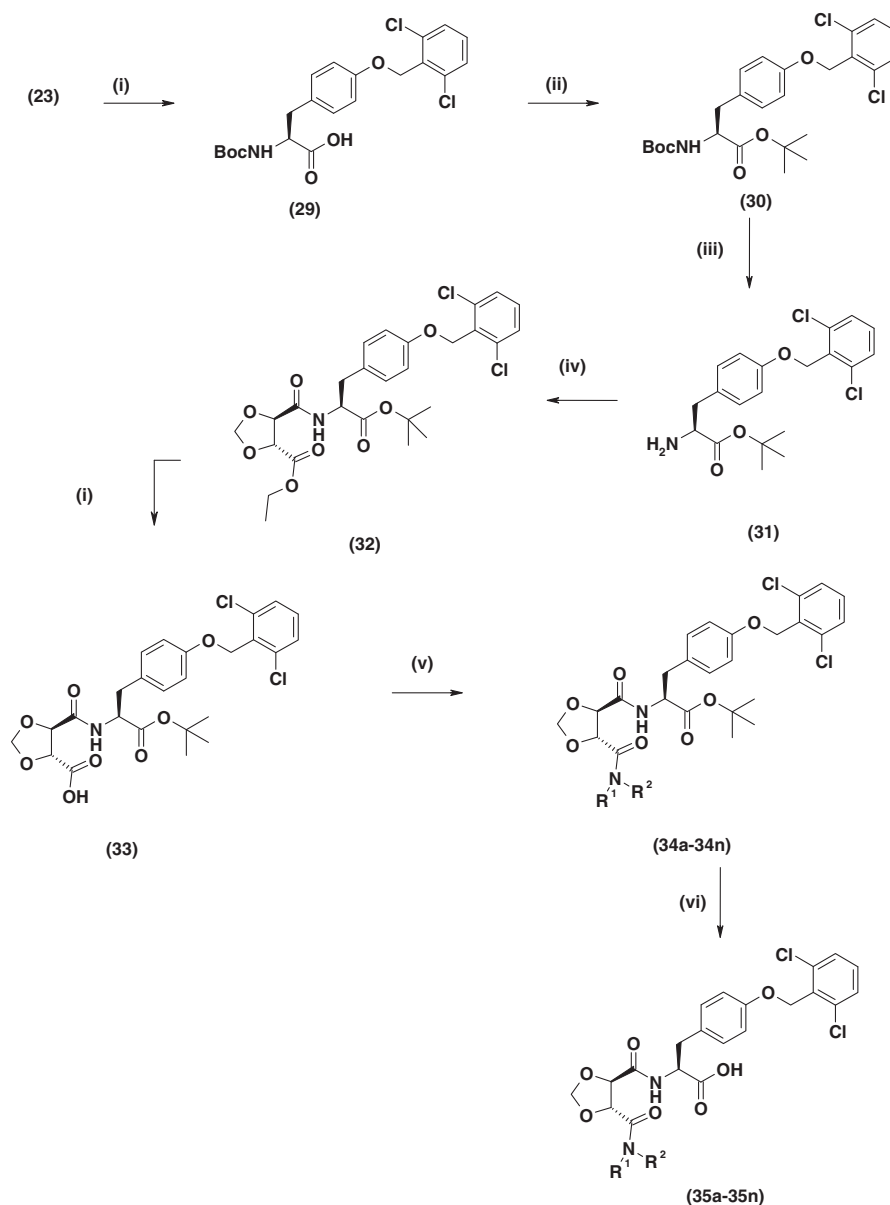
tained an *N*-(2-phenyl)phenylaminocarboxy group attached to the proline ring. We now report the results of further work wherein 1,3-dioxolane ring (8) was evaluated as bioisoster of proline in standard *N*-acylphenylalanine derivatives (7).

(+)-Diethyl-L-tartrate (9) was reacted with dimethoxymethane in the presence of borane trifluoride to form the 1,3-dioxolane derivative (10). Selective hydrolysis of a single ethyl ester group was performed under basic conditions and the resulting mono acid (11) was coupled with 2-phenylalanine using EDC and HOBt. The ester group of the amide (12) (Scheme 1) was hydrolyzed and the acid (13) was coupled with 4-nitro or 4-hydroxyphenyl-L-alanine methyl ester (14a or 14b) to give compounds (15a and 15b). The nitro group of (15a) was reduced by catalytic hydrogenation and the aniline (16) coupled with various acids to give amides (17a–17g). Finally, the esters were hydrolyzed under basic conditions to produce the desired acids (18a–18g). To prepare ether derivatives, the phenolic group (15b) was alkylated with a range of benzyl bromides in the presence of potassium carbonate to furnish the desired esters (19a–19c) which were hydrolyzed under basic conditions to the acids (20a–20c).

In order to provide structure diversity in the acyl portion we prepared a number of amide derivatives by one of two different methods. In the first method (Scheme 2), the amino group of tyrosine (21) was Boc protected and the phenolic group of the product (22) alkylated using 2,6-dichlorobenzyl bromide and potassium carbonate to furnish ether (23). Deprotection of the Boc group under acidic conditions gave amine (24). Intermediate (11) was reacted with various amines to produce amides (25a–25n). Basic hydrolysis of the ester groups gave acids (26a–26n) which were



Scheme 2. Reagents: (i) Boc anhydride, Et₃N, DCM, 100%; (ii) 2,6-dichlorobenzyl bromide, K₂CO₃, acetone, 74%; (iii) TFA, DCM, 100%; (iv) RNH₂, EDC, NMM, HOBt, DMF, 70–80%; (v) LiOH, THF, MeOH, H₂O, 70–80%; (vi) EDC, NMM, HOBt, DMF, 50–75%.



Scheme 3. Reagents and conditions: (i) LiOH, THF, MeOH, H₂O, 90%; (ii) *t*-BuOH, DMAP, rt, 30%; (iii) H₂SO₄, *t*-BuOAc, 78%; (iv) (11) EDC, NMM, HOBt, 68%; (v) R¹R²NH, EDC, NMM, HOBt, DMF, 60–80% (34a–34e) or R¹R²NH, ethylchloroformate, NMM, THF, 70–85% (34f–34n); (vi) TFA, DCM, 80–90%.

coupled with (24) to afford amides (27a–27n). Final hydrolysis gave the target compounds (28a–28n).

In the second method (Scheme 3), the intermediate (23) was hydrolyzed under basic conditions to afford the acid (29) which was protected as a *t*-butyl ester (30). Selective Boc deprotection gave amine (31) which was coupled with intermediate (11) to furnish the amide (32). The methyl ester group was hydrolyzed to afford the acid (33) which was coupled with a diverse set of amines to afford amides (34a–34n). Hydrolysis under acidic conditions then yielded the final compounds (35a–35n).

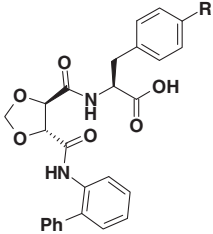
A diverse set of compounds incorporating a 1,3-dioxolane ring in place of the proline ring normally found in many VLA-4 antagonists were synthesized and evaluated as potential VLA-4 antagonists by measuring their ability to inhibit binding of VLA-4 to VCAM-1 in a U-937 (human T-cell line) cell adhesion assay.¹³

In the first part of the work, an exploration was undertaken of the effect on binding to the VCAM-1 receptor of compounds containing various substituents at the *para*-position of the phenyl ring of the phenylalanine group (Table 1). It was observed in the series

of compounds (18a–18g) that 2,6-di-substitution of the phenylcarboxamido group gave a compound with higher binding affinity than the unsubstituted or mono-*ortho*-substituted compound (compare 18e with 18g and 18f). The high binding affinity of the 2,6-dichloro derivative 18e may arise from either an electron-withdrawing or steric effect. However, an examination of the results with the pyridylcarboxamido analogs (18a–18d) indicates that these compounds do not have particularly high binding affinities, thereby implying that the high binding affinity of compound 18e arises from favorable steric and/or hydrophobic interactions at the receptor. A similar trend is observed in the series of derivatives in which an ether group is placed at the *para*-position of the phenylalanine group (20a–20c). The 2,6-dichlorophenylmethylether (20c) has a far higher receptor binding affinity than the 2-chlorophenylmethyl ether (20b) or the 2,6-difluorophenylmethylether (20a), suggesting once again that steric as opposed to electrostatic forces are at work.

In the second part of the work, the favored 2,6-dichlorophenylmethyl substituent was retained at the *para*-position of the

Table 1
Activity of 1,3-dioxolanes as VLA-4 antagonists



Compd	R	IC ₅₀ (μM)	n
18a		8.3	1
18b		2.7 ± 0.46	2
18c		9.5	1
18d		6.3	1
18e		0.1	1
18f		9.0	1
18g		6.3	1
20a		7.9	1
20b		8.7 ± 3.74	2
20c		0.5 ± 0.25	3

phenylalanine group and attention turned to substitution of the 1,3-dioxolane ring in a study of a range of N-substituted aminocarbonyl-1,3-dioxolanes (Table 2). Very little of a structure–activity trend was seen in the N-alkyl amides (**28a–28j**, **35n**) with all compounds having IC₅₀ values in the range of 1–7 μM with the exception of the N-cyclopropyl amide (**28j**) which had an IC₅₀ value of 0.3 μM. In the set of N-aryl amides, *ortho*-substitution consistently gave compounds (**35a** and **35b**, **35d**, **35f**) having higher binding affinities than unsubstituted (**35e**) or *para*-substituted (**35c**) derivatives. An additional beneficial effect on activity was not observed in compounds containing a second *ortho*-substituent (compare the *ortho*-substituted compounds, **35b** and **35a**, with the 2,6-disubstituted compounds, **35j** and **35i**, respectively). Electronic effects do

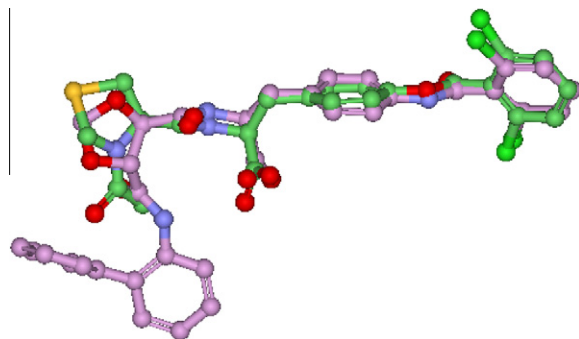


Figure 3a. Structural overlay of CT-5219 (green) with **18e**.

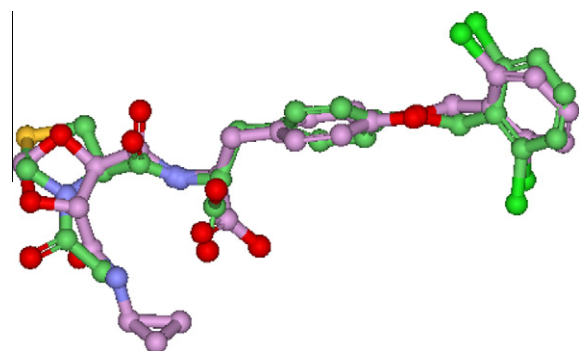


Figure 3b. Structural overlay of CT-5219 (green) with **28j**.

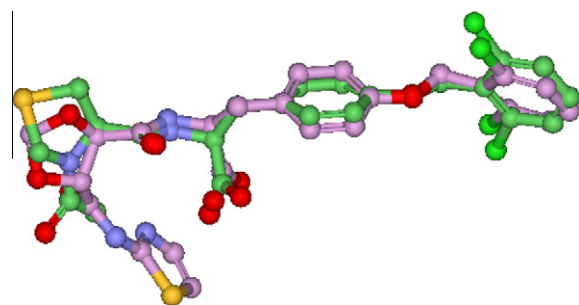
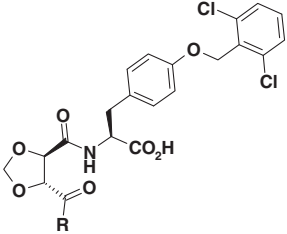


Figure 3c. Structural overlay of CT-5219 (green) with **35g**.

not appear to be playing a major role in this case because the 3,5-dichloropyrid-4-yl derivative (**35k**) had a similar binding affinity to the phenyl analog (**35j**). Compounds containing electron-rich aromatic systems such as thiazol-2-yl (**35g**), benzthiazol-2-yl (**28k**), thien-2-ylmethyl (**28l**), and indol-3-ylethyl (**28m**) all had sub-micromolar affinity which was reduced when the aromatic systems were substituted (**35l** and **35m**, **28n**). It appears, therefore, that a range of substituents are tolerated on the nitrogen atom of the aminocarbonyl-1,3-dioxolane, indicating perhaps that this part of the molecule plays a limited role in the binding of the ligands to the VCAM-1 receptor. When tested for metabolic stability in human liver microsomes (HLM) at 8 μM concentration for 1 h, compounds (**28j**) and (**35g**) were found to be stable 72% and 52%, respectively.

In conclusion, we have successfully demonstrated that the proline ring of the N-acylphenylalanine class of VLA-4 antagonists can be replaced by a range of N-substituted aminocarbonyl-1,3-dioxolanes. Three compounds with high affinity for the VLA-4 receptor, each having a 2,6-dichlorophenyl substituent attached by means of an amide or ether linkage to the *para* position of the phenylalanine group, were the N-(2-phenyl)phenyl amide (**18e**), N-cyclopro-

Table 2
Activity of 1,3-dioxolanes as VLA-4 antagonists



Compd	R	IC ₅₀ (μM)	n	Compd	R	IC ₅₀ (μM)	n
28a		1.4 ± 0.55	2	35a		0.6 ± 0.25	2
28b		1.2 ± 0.75	3	35b		0.7 ± 0.25	3
28c		2.1 ± 1.06	2	35c		2.8	1
28d		1.5 ± 0.40	2	35d		0.6 ± 0.2	3
28e		1.1 ± 0.45	3	35e		1.5 ± 0.7	2
28f		4.4 ± 3.4	2	35f		0.9 ± 0.5	3
28g		1.5	1	35g		0.2 ± 0.1	3
28h		6.6	1	35h		1.5	1
28i		1.2 ± 0.1	3	35i		0.5 ± 0.1	2
28j		0.3 ± 0.1	3	35j		0.7 ± 0.3	3
28k		0.6 ± 0.01	1	35k		0.4	1
28l		0.7 ± 0.3	3	35l		1	1
28m		0.6 ± 0.2	3	35m		2.6	1
28n		3 ± 0.3	2	35n		3.4	1

pyl amide (**28j**), and *N*-(thiazol-2-yl) amide (**35g**). Interestingly, overlay of the three compounds indicates that orientation of the functional groups of these compounds were similar to **CT-5219**

in the energy minimized structure (Figs. 3a–c). Subsequent work on the further optimization of this series of compounds will be disclosed in due course.

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